Clostridium difficile Infection:

Energing
Eherapies
and Recurrence
Mrevention

Internet Release Date:

December 20, 2011

ACPE Live Release Date:

October 22, 2011

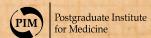
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RMEI gratefully acknowledges an educational grant from Optimer in support of this CE activity



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WELCOME

Thank you for joining us for *Clostridium difficile* Infection: Emerging Therapies and Recurrence Prevention, a continuing education activity.

We also thank our esteemed speakers for sharing their time and expertise. Through this activity, they will describe the role of gastrointestinal flora in health and the mechanisms of recurrent *Clostridium difficile* infection (CDI); identify patients at risk for recurrence and poor outcomes; summarize recent evolutions in CDI treatment options; use appropriate CDI diagnostic testing; and choose appropriate infection control methods and therapies to improve patient outcomes.

This workbook includes the presenters' slides and will help guide you through the activity. If you would like to receive 2.0 *AMA PRA Category 1 Credits* $^{\text{TM}}$, 2.0 ACPE contact hours, or 1.9 ANCC contact hours, please complete the activity evaluation as instructed on this activity's website.

We hope that you will find this activity rewarding and informative.

AGENDA

Activity Overview Dale N. Gerding, MD, FACP, FIDSA Diagnostic Challenges and Controversies: Becoming Less Difficile Stephen M. Brecher, PhD Clostridium difficile: Pathophysiology, Risk Factors, and Recurrence Kathleen Mullane, DO, PharmD **Emerging CDI Treatment Options and Infection Control** Stuart Johnson, MD, DTM&H **Question-and-Answer Session**

OVERVIEW

Target Audience

This activity has been designed to meet the educational needs of physicians, pharmacists, and nurses involved in the care of patients with *Clostridium difficile* infection (CDI).

Statement of Need

Rates of CDI incidence, hospitalizations, and deaths have been increasing in recent years, suggesting that current management of this condition may be suboptimal.¹ Although much has been discovered regarding the role of gastrointestinal flora in the pathogenesis of CDI, the mechanisms of recurrent disease and poor outcomes remain elusive. Surveys of practice patterns suggest that healthcare professionals are unaware of or fail to utilize optimal methods for managing patients with CDI.^{2,3} Given the recent updated guidelines for CDI, combined with several new agents emerging for this disease, it is important that healthcare professionals be aware of the most current research and recommendations for the management of CDI.

1 Zilberberg MD, et al. Emerg Infect Dis. 2008;14:929-931.

2 Byker GL, et al. Infect Control Hosp Epidemiol. 2009;30:397-399.

3 Association for Professionals in Infection Control (APIC) Pace of Progress poll. May 2010.

Educational Objectives

After participating in this educational activity, participants should be able to:

- Describe the role of gastrointestinal flora in health and the mechanisms of recurrent Clostridium difficile infection (CDI)
- Identify patients at risk for recurrence and poor outcomes
- Summarize recent evolutions in CDI treatment options
- Use appropriate CDI diagnostic testing
- Choose appropriate infection control methods and therapies to improve patient outcomes

Statement of Support

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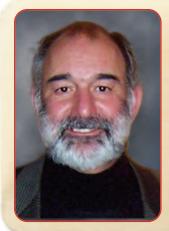
Dale N. Gerding,
MD, FACP, FIDSA (Moderator)

Professor of Medicine
Loyola University Chicago
Stritch School of Medicine
Maywood, IL
Research Physician
Hines Veterans Affairs Hospital
Hines, IL

Dale N. Gerding, MD, FIDSA, is Professor of Medicine at Loyola University Chicago Stritch School of Medicine in Maywood, Illinois and Research Physician at Hines Veterans Affairs Hospital. Prior to his present position, Dr. Gerding was Associate Chief of Staff for Research and Development at the Hines Veterans Affairs Hospital, Chief of Medicine at Lakeside VA Hospital in Chicago, and Professor of Medicine at Northwestern University Feinberg School of Medicine. Dr. Gerding received his undergraduate degree in physics from St. John's University in Collegeville, Minnesota, attended graduate school in physics at UCLA in Los Angeles, California, and received his MD from the University of Minnesota Medical School. He was a medical intern at the Peter Bent Brigham Hospital in Boston and, following two years at the National Institutes of Health, completed his medical residency and infectious diseases fellowship at the University of Minnesota and Minneapolis VA Medical Center. He is board certified in internal medicine and infectious diseases.

Dr. Gerding is an infectious diseases specialist and hospital epidemiologist, past president of the Society for Healthcare Epidemiology of America and past chair the antibiotic resistance committee of SHEA. He is a fellow of the Infectious Diseases Society of America and past chair of the National and Global Public Health Committee and the Antibiotic Resistance Subcommittee of IDSA. He served as a member of the board of directors of IDSA from 2005-2008. He is a fellow of the American College of Physicians and a member of the American Society for Microbiology.

Dr. Gerding's research interests include the epidemiology and prevention of *Clostridium difficile* disease, antimicrobial resistance, and antimicrobial distribution and kinetics. He has been a Merit Review funded research investigator in the VA for over 35 years and is the author of over 300 peer-reviewed journal publications, book chapters, and review articles. He holds patents for the use of non-toxigenic *C. difficile* for the prevention and treatment of this disease. He is a member of the editorial boards of *Clinical Infectious Diseases*, *Antimicrobial Agents and Chemotherapy*, *Gut Microbes*, and *Infection Control and Hospital Epidemiology*, and is an ad hoc reviewer for numerous other medical journals.



Stephen M. Brecher, PhD

Director of Microbiology VA Boston Healthcare System West Roxbury, MA

Dr. Stephen M. Brecher has been the Director of Microbiology at the Boston VA Healthcare System for 28 years and holds academic appointments at the Boston University School of Medicine and at the University of Massachusetts/Dartmouth. Dr. Brecher is on the editorial board of the *Journal of Clinical Microbiology*.

Dr. Brecher has been honored as an American Society of Microbiology (ASM) Foundation Speaker and has convened and lectured at numerous symposia and workshops at the ASM annual meetings (the general and "infections meetings"). Dr. Brecher is well known and respected for his work and lectures on methicillin-resistant *S. aureus* (MRSA), *C. difficile* infections (CDI), and multi-drug resistant bacteria. He has lectured from Boston to Beijing and, as an accomplished and highly acclaimed speaker, Dr. Brecher is appreciated for his ability to make you laugh while he is telling you that bacteria are the dominant species on earth and that your days are numbered. CD infections are particularly important to him because of the poor outcomes, treatment difficulties, diagnostic dilemmas, and unexplained community cases.



Kathleen Mullane,

Associate Professor of Medicine University of Chicago Chicago, IL

Kathleen M. Mullane, DO, PharmD, is Associate Professor of Medicine at the University of Chicago in Chicago, Illinois. Following graduation from the Chicago College of Osteopathic Medicine at Midwestern University, Dr. Mullane completed her residency in internal medicine at Rush-Presbyterian-St. Luke's Medical Center and her fellowship in infectious diseases at the University of Illinois/University of Chicago Combined Program.

Dr. Mullane's clinical interest is in infectious diseases and antimicrobials. She has co-authored numerous original articles and abstracts on investigational treatments for infectious diseases including human immunodeficiency virus (HIV) and hepatitis C, as well treatment of skin, soft tissue, and fungal infections. Dr. Mullane has also participated in numerous lectures and grand rounds on topics such as HIV, West Nile virus, tuberculosis, fungal infections, and sexually transmitted diseases.

Dr. Mullane currently holds positions on the *American Journal of Transplantation* and *Transplant Infectious Diseases* journal review panels. She is the Director of Infectious Diseases Clinical Trials, Chairman of the Antibiotic Subcommittee, and a member of various BSD committees at the University of Chicago. In 2004, she was the recipient of the Inspirational Attending of the Year at Loyola University Medical Center.



Stuart Johnson,
MD, DTMS-H

Professor of Medicine
Loyola University Medical Center
Stritch School of Medicine
Maywood, IL
Staff Physician
Hines VA Hospital
Hines, IL

Stuart Johnson, MD, DTM&H, is a professor of medicine at Loyola University Stritch School of Medicine at Loyola University Medical Center in Maywood, Illinois and the Deputy ACOS for Research at the Hines VA Hospital. Dr. Johnson received his MD from the University of Minnesota Medical School and completed his internal medicine residency and infectious disease fellowship training at the University of Minnesota Hospital and the Minneapolis VA Medical Center. He received a diploma in Tropical Medicine and Hygiene at Mahidol University in Bangkok, Thailand and a Career Development Award from the Department of Veterans Affairs. He is the past president of the Anaerobe Society of the Americas.

His main research interest and focus has involved the epidemiology and pathogenesis of *Clostridium difficile* infections. He is actively studying variant strains of *C. difficile* and the role of the various toxins in the pathogenesis of *C. difficile* disease. He also has been involved in clinical research on the parasite, *Angiostrongylus cantonensis*, responsible for most cases of eosinophilic meningitis, world-wide.

ACCREDITATION & CREDIT

Physician Continuing Education

Accreditation Statement

This activity has been planned and implemented in accordance with the Essential Areas and policies of the Accreditation Council for Continuing Medical Education through the joint sponsorship of Postgraduate Institute for Medicine and RMEI, LLC. Postgraduate Institute for Medicine is accredited by the ACCME to provide continuing medical education for physicians.

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Credit Designation

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Live ACPE Release Date: October 22, 2011

Type of Activity: Knowledge

Pharmacy Educational Objective

After completing this activity, the participant should be able to:

Provide accurate and appropriate counsel as part of the treatment team

A statement of credit will be issued only upon receipt of a completed activity evaluation form and will be mailed to you within 3 weeks.

Nursing Continuing Education

Credit Designation

This educational activity for 1.9 contact hours is provided by Postgraduate Institute for Medicine.

Accreditation Statement



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California Board of Registered Nursing

Provider approved by the California Board of Registered Nursing, Provider Number 13485, for 1.9 contact hours.

A statement of credit will be issued only upon receipt of a completed activity evaluation form and will be mailed to you within 4 weeks.

Nursing Educational Objective

After completing this activity, the participant should be able to:

• Provide appropriate care and counsel for patients and their families

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There are no fees for participating and receiving CE credit for this activity. During the period December 20, 2011 through December 20, 2012, participants must read the learning objectives and faculty disclosures and study the educational activity.

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Media

Internet

Fee Information

There is no fee for this educational activity.

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- Dale Gerding, MD, FACP, FIDSA, has affiliations with ViroPharma (*Patent*); ViroPharma, Merck, Pfizer, Optimer, Cubist, TehraDoc, Medicines Co., Astellas, and Actelion (*Advisory Board*); and Merck, Optimer, GOJO, Sanofi Pasteur, ViroPharma, Actelion, and Eurofins Medinet (*Research*).
- Stephen M. Brecher, PhD, has no affiliations with commercial interests to disclose.
- Kathleen Mullane, DO, PharmD, has affiliations with Actelion Pharmaceuticals Ltd, Cubist, Sanofi Pasteur SA, and ViroPharma Incorporated (*Research*); and Merck & Co., Inc., Novartis AG, and Optimer Pharmaceuticals, Inc. (*Research and Consultant*).
- Stuart Johnson, MD, DTM&H, has affiliations with Optimer Pharmaceuticals, Inc., ViroPharma Incorporated, Astellas Pharma US, Inc., Pfizer, Cubist Pharmaceuticals, and Bio-K+ (Consultant).

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- Nora Duffy has no affiliations with commercial interests to disclose.

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- Jan Hixon, RN, BSN, MA, has no affiliations with commercial interests to disclose.
- Trace Hutchison, PharmD, has no affiliations with commercial interests to disclose.
- Julia Kimball, RN, BSN, has no affiliations with commercial interests to disclose.
- Samantha Mattiucci, PharmD, has no affiliations with commercial interests to disclose.
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Disclaimer

Participants have an implied responsibility to use the newly acquired information to enhance patient outcomes and their own professional development. The information presented in this activity is not meant to serve as a guideline for patient management. Any procedures, medications, or other courses of diagnosis or treatment discussed or suggested in this activity should not be used by clinicians without evaluation of their patient's conditions and possible contraindications on dangers in use, review of any applicable manufacturer's product information, and comparison with recommendations of other authorities.



1

Diagnostic Challenges and Controversies:
Becoming Less Difficile

Stephen M. Brecher, Ph.D.
VA Boston HealthCare System
BU School of Medicine

The opinions expressed in this presentation are those of the presenter and do not necessarily represent the views of the Veterans Affairs Health-Care System

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Overview

- · Recent guidelines for testing
- Overview of testing
- · RIP for EIA
- Molecular tests
- · Repeat testing
- Mural dyslexia

The Most Accurate Method to Diagnose C. difficile is

- 1. EIA
- 2. GDH
- 3. EIA/GDH combined
- 4. PCR

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Specimens for *C. difficile* Should Be

- 1. Frozen before testing to release toxin
- 2. Heated to select for spores
- 3. Stored at room temperature
- 4. Fresh and loose

What Do You Want for CDI Testing?

- 1. An assay with low sensitivity and high specificity (false negatives)
- 2. An assay with high sensitivity and low specificity (false positives)
- 3. An assay that is inexpensive, easy, and quick to perform
- 4. I want to get it right even if it is very expensive

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Clinical Practice Guidelines 2010 SHEA and IDSA¹ Summary

- · Test only unformed stool (exception: ileus)
- · Do not perform a test of cure
- Stool cultures sensitive but not practical except for epidemiological studies
- · EIA is rapid, not very sensitive and is sub-optimal
- · 2 step GDH and EIA is an interim recommendation
- More data needed on PCR before they can recommend
- Repeat testing discouraged

Cohen SH, et al. Infection Control and Hospital Epidemiology, 2010;31:431-455

A Practical Guidance Document for the Laboratory Detection of Toxigenic Clostridium difficile ASM September 21, 2010 (ASM online)

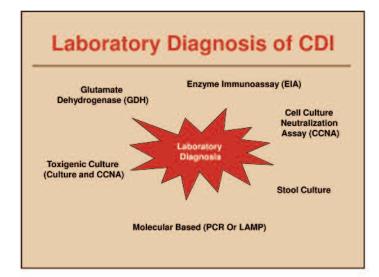
- Utilizing toxin A/B EIA for C. difficile diagnosis is insensitive and no longer recommended as a standalone test
- GDH antigen assays have been found to be good screening tests for CDI in many studies (high sensitivity and NPV)
- +GDH must be confirmed (need + A/B EIA or + cytotoxin or + nucleic acid amplification test (NAAT))
- · NAAT can be used as a stand alone test
- · Do not perform a test of cure

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C. difficile Testing 1984 – 2010 Ducks in a Row?



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Specimen and Testing Guidelines

- · Use the "Brecher" guidelines
 - Only test loose or liquid stool
 - · "If it ain't loose, it's of no use"
 - Stick test for stool consistency
 - "If the stick stands, the tests are banned if the stick falls, test them all"
- · Limit testing to 1 test/patient/week (if using PCR)
- · Do not perform a test for cure
- · Do not perform tests on asymptomatic patients

CDI Testing Issues

- · What is the gold standard?
- · Is it time to abandon EIA?
- What about 2-3 step algorithms (difficile dancing)?
- · Is PCR/molecular ready for prime time?

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The Gold Standard is Tarnished*

- All C. difficile test assay studies are flawed
- There is no reliable gold standard¹
- · Patients can carry toxigenic strains
- Suggested gold standard has to include a very reliable assay as well as the clinical status of the patient

"Plate Sin With Gold and it Goes Untarnished" W. Shakespeare

1. Wilcox MH, et al. J Clin Microbiol. 2010;48(12):4347-4353.

CDI Testing Issues

- · What is the gold standard?
- · Is it time to abandon EIA?
- · What about 2-3 step algorithms (difficile dancing)?
- · Is PC

15

R/molecular ready for prime time?	
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Conflicting Results with EIA

Recently Published EIA Papers(1-6)			
Parameter	Range		
Sensitivity	32 – 98.7%		
Specificity	92 – 100%		
PPV	76.4 – 96%		
NPV	88 – 100%		

With an average sensitivity of 60-70%, do not perform as a stand-

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CDI Testing Issues

- What is the gold standard?
- Is it time to abandon EIA?
- · What about 2-3 step algorithms (difficile dancing)?

Is PCR/molecular ready for prime time?

C. Diff Quik Chek Complete

- · Lateral flow GDH and EIA on one test card
- 2 recent studies
- Quinn et al1 reported that if
 - Both + = +
 - . Both = -
 - 13.2% discrepant, re-test. Use PCR
 - Sharp et al2 reported that 88% of specimens were both positive or both negative
 - · Used random access PCR to resolve remaining 12%
- Quinn CD, et al. J Clin Microbiol. 2010;48:603-605. Sharp SE, et al. J Clin Microbiol. 2010;48:2082-2086

Possible Explanation for EIA and GDH Tests Failure

- Tenover et al¹ reported that EIA testing of certain ribotypes (002, 027, and 106) had very poor sensitivity compared to PCR
 - EIA test failures may be associated with geographical ribotype differences
- GDH had lower sensitivity for non-027 ribotypes
- GDH has equal sensitivity (to PCR) for 027 ribotypes

1. Tenover FC, et al. J Clin Microbiol. 2010;48:3719-3724.

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CDI Testing Issues

- · What is the gold standard?
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- What about 2-3 step algorithms (difficile dancing)?
- Is PCR/molecular ready for prime time?

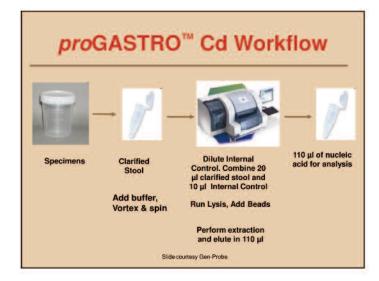
Commercially Available Molecular Assays

Assay	Target	Instrument	TAT
BD GeneOhm™ Cdiff Assay	ToxinB	SmartCycler Amplification	75-120 min
Prodesse proGASTRO™	Toxin B	easyMag extraction SmartCycler Amplification	~ 3 hrs
Cepheid Xpert™ C. difficile	Toxin B+/- tcdC deletion	GeneXpert	45 min
Meridian Illumigene	conserved 5' sequence of the tcdA gene	illumigene Incubator/reader	45-60 min

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BD GeneOhm™ C. diff Assay

	-			16	
		10	O		NO STATE OF THE PARTY STATE OF T
Stool Specimen	Specimen Preparation	Lysis - DNA Extraction	Reconstitution Of Reagents	Real-time PCR Analysis on the SmartCycler*	Definitive On-action Results
10 11 1	1 2 3		sults in <	2 Hours	

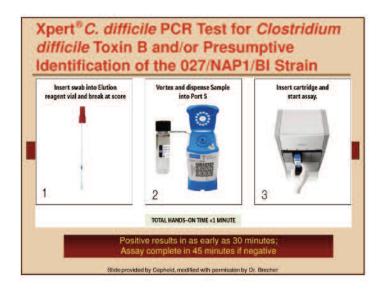


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Program instrument and run assay

Hands-on Time = 57 min
Hands-off Time = 137 min

Slide courtesy Gen-Probe



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- 11 minute sample extraction with -2 minutes of hands on time.

- No centrifugation required.

- All required material included in kit.

- Identical simple process for single or batched samples.

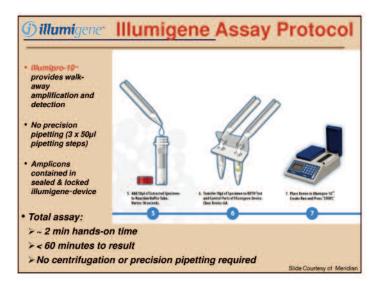
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Summary of C. difficile PCR Published Data (2010-2011)

Publication	PCR Assay	Sens/Spec
Chapin, 2011 ⁷	Compared 3 molecular methods	88.5-96.2%/91.6-100% (range)
Noren, 2011 ²	Illumigene (LAMP)	98%/98.%
Kvach, 2010 ³	BD GeneOhm	91.4%/100%
Novak-Weekley, 2010 ⁴	Cepheid Xpert	94.4%/96.3%
Swindells, 2010 ⁵	Cepheid Xpert BD GeneOhm	100%/99.2% 94.4%/99.2%

- Chapin KC, et al. JMD. 2011;13:395-400.

 Noren T. et al. J Clin Microbiol. 2011;49:710-711.

 Kvach EJ, et al. J Clin Microbiol. 2010;48:109-114.

 Novak-Weckley, et al. J Clin Microbiol. 2010;48:808-803.

 Swindells J, et al. J Clin Microbiol. 2010;48:806-608.

2	8

Diagnostic Accuracy of Real-time Polymerase Chain Reaction in Detection of *Clostridium difficile* in the Stool Samples of Patients With Suspected *Clostridium difficile* Infection: A Meta-Analysis

- •19 studies, 7392 samples, 15 years (1995-2010)
- •Mean sensitivity was 90%
- •Mean specificity was 96%
- ·As prevalence increased, accuracy increased

Deshpande A. et al. Clin Infect Discord. 2011;53:e81-e90. October, 2011.

Camp Clin Micro

- · 40 Microbiologists met in Houston last February
- · Consensus opinion on C. difficile testing1
 - EIA for Toxins A&B should not be used
 - 2 step tests (GDH and Toxins A&B) with confirmation of discrepant results acceptable
 - Used appropriately, molecular based tests are preferred

1. Carroll KC, et al. J Clin Microbiol. 2011;49:S49-S52.

If the First PCR is Negative Should I Order Another PCR?

- Of 406 tests from 293 patients with a prior negative PCR¹
 - -396 negative
 - 10 positive
 - · Only 3+ in <7 days
- Exceptions
 - Severe clinical changes

1. Luo RF, Banaei N. J Clin Microbiol. 2010;48:3738-3742

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Consequences of Unreliable CDI Assays

- · Repeat testing (common to order x 3)
- · Low sensitivity
 - False negative patients don't get treated and spread the organism¹
- Low specificity
 - False positive patients get costly treatments and IC protocols

1. Guerrero DM, et al. Clin linfect Dis. 2011;53:287-290.

Consequences of Improved CDI Assays

- · Increased sensitivity
 - True prevalence (will go up with PCR)
 - Infected patients treated and put on Infection Control (IC) protocols sooner
- · Increased specificity
 - Prevent unnecessary treatment
 - Improved utilization of IC protocols
- · Increased productivity
 - Eliminate duplicate/triplicate testing
 - Improves lab utilization and image

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Which Molecular Test Should I Use?

- · Do your homework
- · Look at
 - Reagent costs
 - Equipment costs
 - TAT/Hands on time
 - Batch or on demand
 - Prevalence of CDI
 - Cost of incorrect tests

Do I Need to Test for the "Hypervirulent" Ribotype?

- The fidaxomycin treatment relapse rate was significantly lower than the vancomycin relapse rate for all strains except NAP1/B1/027¹
- The Cepheid PCR assay can detect the gene deletion associated with this strain (tcdC)
- Recommend treating patients based on symptoms and severity of disease, not the strain
 - Need more data

1. Louie TJ, et al. N Eng J Med. 2011;364:422-431

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Mural Dyslexia

- We don't see the big picture because we have become bean counters
 - "we can't afford to do PCR"
- We can save money by spending money
- With an accurate method and strict specimen requirements, test volume will decrease
 - Boston VA HCS: Test volume decreased by 50% with PCR
- · With the correct diagnosis
 - Appropriate treatment gets patients home faster and that's the way to save money (LOS)
 - IC dollars spent on the right patients

Recommendations

- · Test by a molecular method
- Test only unformed stool in symptomatic at-risk patients
- Test only 1 stool/patient/week
- · Do not perform a test of cure
- Remember no lab test is perfect so correlate test results with patient data/clinical observations
- · There is light at the end of the colon!!!

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C. difficile Testing 2011 Heading in the Right Direction



Clostridium difficile:

Pathophysiology, Risk Factors, and Recurrence

Kathleen M. Mullane, D.O., Pharm. D., FIDSA University of Chicago Department of Medicine / Section of Infectious Diseases 1

Disclosures

- · Clinical trials research in CDI (C. difficile infection)
 - Actelion
 - Cubist
 - Merck
 - Novartis
 - Optimer
 - Sanofi-Pasteur
 - Viropharma
- · Protocol / publication committees
 - Merck
 - Novartis
 - Optimer

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4

CDI Pathophysiology

- · Ingestion of vegetative organisms and spores
- · Overgrowth of toxigenic strains of C. difficile
 - Antibiotics
 - Chemotherapy
- · Production of toxins
 - · Toxin A (TcdA) classically: endotoxin
 - · Toxin B (TcdB) classically: cytotoxin
 - · C. difficile binary toxin (CDT) some strains
 - · Isolates lacking toxins are nonpathogenic

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CDI Pathophysiology

- · Development of a range of clinical signs
 - Mild to severe self-limiting diarrhea
 - Life-threatening pseudomembranous colitis
 - Toxic megacolon / intestinal perforation
 - Systemic inflammatory response syndrome (SIRS)
 - · Intestinal tissue damage and inflammation

Patient Characteristics as Risk Factors for Acquisition of CDI

- · Aged > 65 years
- · Multiple Medical Co-morbidities
 - Antibiotic therapy
 - · Cephalosporins, Clindamycin, Fluoroquinolones
 - Inflammatory bowel disease
 - Chemotherapy / Bone marrow transplant
 - Renal impairment
 - Human immunodeficiency virus (HIV)
 - Organ transplant
- · PPI

2008 Nationwide Inpatient Sample, 2009 Medicare Provider Analysis and Review file, Florida fiscal year 2005-2007 Medicald claims and 2007-2008 Medicare 5% Standard Analytical Files. Dubbarke, et. al. SHEA April 2, 2011: Dallas T.

Predictors of H	ospital-Acq	uired CDI
-----------------	-------------	-----------

- Hospital-level predictors
 - More likely to have concurrent infections (rank)
 - · Pseudomonas, Staphylococcus, Streptococcus, E. coli
 - Discharge diagnosis of infection
 - Bloodstream infections, endocarditis, osteomyelitis, pneumonia, UTI and cellulitis
 - Low nurse : patient and resident : patient ratio
 - Slow bed turnover (ie: longer LOS)
 - · Cardiac intensive care services
 - · Transplant services

Ricciard R. Epidemiol Infect. 2008;136(7):913-92

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Risk Factors for Community Acquired CDI

- Antimicrobial drug exposure ≤ 1 month
- · Anti-motility agent use
- Recent hospitalization (< 6 months)
- · Markers of chronic disease
 - Frequent outpatient visits
 - GERD (? Proton pump inhibitors / H2 blockers)
 - · OTC use of agents may not be well accounted
 - Cardiac failure
- Contact with infants ≤ 2 years

Wilcox MH, et al. J Antimicrob Chemother, 2008;62:388–396 Wilcox MH, et al. Arch Int Med. 2010;170(9):785-790. Kutty PK, et al. Emerg infect. Dis. 2010;16(2):197-204.

Communit	y Acc	uired	CDI	Risk
	Fact	ors		

- Foodborne: Meats and produce?
 - C. difficile isolated from the feces of food animals
 - · Cattle/calves, pigs/piglets, horses and chickens
 - Role in causing disease in some animals is unclear, but is well documented in piglets (typhylocolitis)
 - Few small studies in various geographic regions with different sampling / culture methods
 - · Colonizaton rates decrease from sucklings to finishers
 - Potential concern: shedding by asymptomatic animals
 causing contamination of food, water, and the environment
 - · Etiology of colonization unknown (humans : animals)
 - PCR Ribotype 078 most common pig strain and increasingly prevalent human strain

Eur Soc Clin Micro Inf Dis 2009; 16: 3-10. Gould LH. Limbago B. Clin Infect Dis. 2010;51(5):577–582. Keessen EC, et al. Vet Microbiol. 2011 Mar 26. [Epub ahead of print

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TcdA and TcdB Multifactorial Activities

- Clostridial Glucosylating toxin family
 - Large Clostridial Toxins (large molecular mass)
 - Direct toxin target is colonic epithelium
 - Receptor mediated endocytosis followed by pore formation and translocation into the cytosol
 - · Target /modify small GTPases of the Rho/Ras family
 - Molecular switches involved in signaling processes
 - » Regulation of the actin cytoskeleton dynamics (Depolymerization disrupts tight junctions breaching intestinal-epithelial barrier / increased permeability)
 - » Gene transcription / expression
 - » Cell cycle progression / cell proliferation
 - » Apoptosis

Sun X , et al. Toxins. 2010;2(7):1848-1880. Giesemann T, et al. J Med Microbiol. 2008;57(Pt 6):690-696

TcdA and TcdB Multifactorial Activities

- Enhanced permeability leads to immune cell stimulation
 - Cytokine / pro-inflammatory cytokine release
 - Mucosal damage causes expression of leukocyte and endothelial adhesion molecules
 - » Neutrophil attachment and migration into mucosa
 - » Acute / intense inflammation with neutrophil infiltration and destruction of enterocytes and colonocytes
 - » Edema and cell damage
- Induction of apoptosis of intestinal epithelial cells (lamina propria and submucosa)
 - · TcdA: monocytes / mitochondria and T cells
 - TcdB : direct mitochondrial apoptosis
- · Necrosis : epithelial necrotic death

Sun X , et al. Toxins. 2010;2(7):1848-1880.

10		

Binary Toxin

- · CDT (C. difficile transferase)
 - Pathophysiologic role not well understood
 - Less than 10% of C. difficile isolates
 - Including 100% of NAP1/027 (toxinotype III)
 - Actin-ADP-ribosylating toxin
 - · Cytopathic in cell culture
 - Modifies G-actin
 - · Induces formation of microtubule-based protrusion
 - Increases adherence of bacteria to intestinal epithelium

П	1
-	-

Risk factors for Severe CDI

- Age >70
- · Leukocyte count >20,000 cells/ml
- · Albumin <2.5 g/dl
- · Creatinine >2 mg/dl
- Presence of small bowel obstruction or ileus
- Computed tomography showing colorectal bowel wall thickening, stranding, ascites

lenrich TJ, et al. Emerg Inlect Dis. 2009;15(3):415-423

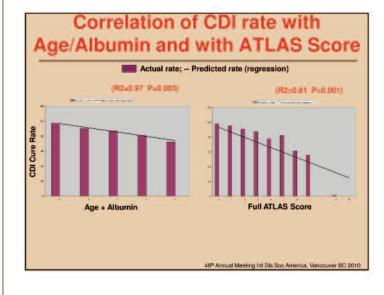
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Prediction of CDI Outcome; ATLAS Scoring System

Parameter	0 points	1 point	2 points
Age (yrs)	< 60	60-79	>80
Temperature (C)	≤ 37.5	37.6-38.5	≥ 38.6
Leukocytosis	<16K	16K-25K	>25K
Albumin (g/L)	>35	26-35	<25
Systemic concomitant antibiotics during CDI therapy	No		Yes

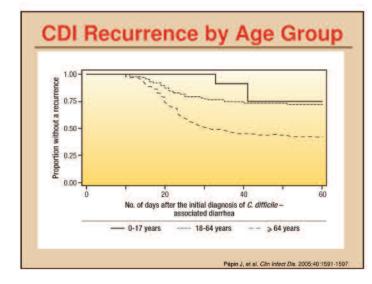
- ATLAS scores for 516 patients with CDI were calculated at their time of diagnosis and matched against their cure rates following 10 days of study treatment
- · Patients with an ATLAS score of 0 had a 98% cure rate
- ATLAS scores of 7 corresponded to a 55% cure rate
 48th Annual Meeting Inf Dis Soc America, Vancouver BC 2010.

13

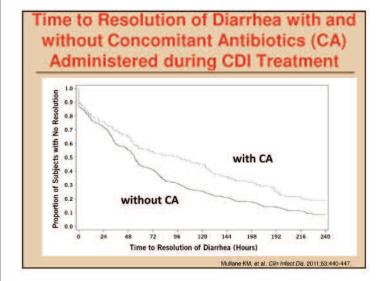


Risk factors for recurrence

- · Old age
- · Concomitant antibiotics
 - Especially fluoroquinolones
- · Low serum albumin
- · Poor performance status
- · Poor immune response against Toxin A or Toxin B
- · Concomitant treatment with antacids / PPIs
- · Concomitant VRE colonization
- · Hospital acquired disease
- · History of surgery
- Fecal incontinence
- Fecal microbiome dysruption



16



17

Effect of Concomitant Antibiotics taken During Treatment or Any Time

(Treatment or follow up)

- · CAs concurrent with CDI treatment:
 - Lower cure rates compared to no CA (84% vs 93%; P < .001)
 - Cure rates for different treatment groups:
 - Fidaxomicin, 90% vs. vancomycin, 79% (P = .04)
- CA administration at any time
 - Lower global cure rate compared to no CA (66% vs 75%; P= .005)
 - Recurrence rates for different treatment groups:
 - Fidaxomicin,16.9% and vancomycin, 29.2%; (P = .048)

Mullane KM, et al. Clin Infect Dis. 2011;53:440-447

18

Clostridium difficile Infection: Emerging Therapies and Recurrence Prevention

Prolonged Hospitalization or Stay in LTCF

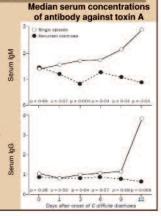
- · Risk related to transmission of C. difficile spores
 - Primary source healthcare workers
 - · may carry C. difficile spores on their hands (not likely fecal carriers)
 - Environmental contamination secondary source
- Up to 50% of LTCF residents and 40% of hospitalized patients have been found to be colonized with C. difficile or its toxin

McFarland LV, et al. N Engl J Med. 1989;320:204-210. Bartlett JG, Gerding DN. Clir Infect Dis. 2006;46(Suppl 1): S12-S18. Simor AE, et al. Infect Control Hosp Epidemol 2002;23:686-703. Hookman P, Barkin JS. World J Gastroenterol. 2009;15:1554-1580.

Defective immune response to toxin A

- Generation of an antibody response to toxin A is associated with protection against symptomatic disease and asymptomatic carriage of C. difficile
- Following symptomatic infection, many individuals develop anti-toxin A and B antibodies
- Inability to acquire immunity to toxin A increases risk for recurrent disease
 - Individuals with recurrent CDI mount poor anti-toxin responses

Giannasca PJ, Warny M. Vaccine. 2004;22:848-856. Kyne L, et al. Lancet. 2001;357:189-193; with permiss



Human Fecal Microbiota and CDI

- Complex ecosystem in symbiosis with host that is altered with antibiotics
 - Epithelium : immune cells : microbiota
 - · 100 trillion bacteria colonize the gut
 - 200 to 1000 distinct bacterial species
 - · All 3 play a defensive role
 - Essential for immune cell development and function
 - » Peyer's patches, lymph nodes, spleen, immunoglobulins
 - Gut microbes cross talk across intestinal epithelium and the immune system
 - · Colonization resistance (competition for space and nutrients)
 - · Products of bacterial metabolism bactericidal
 - Short chain fatty acids, reactive oxygen species
 - Bacteriocidin molecules

Chang JY, et al. J Infect Dis. 2008;197(3):435-438. Jemberg C, et al. 2010;156(Pt 11):3216-3223. Wardwell LH, et al. Curr Infect Dis Rep. 2011;13(1):28-34.

21

22

Intestinal Microbiota & Risk of CDI

- Antibiotic pertubation manifests as decreased colonization resistance of commensals
 - Short term: quantity and composition
 - May have a long term influence on return of gut flora to pre-treatment levels
 - · Alter microbial diversity (Bacteroidetes / Firmicutes)
 - Antibiotic associated diarrhea
 - C. difficile, K. oxytoca, C. perfringens, Salmonella spp., Candida spp., and S. aureus

Wardwell LH, et al. Curr Infect Dis Rep. 2011;13(1):28-34

Recurrent C. difficile Infection

- · Rates of recurrent CDI
 - 20% after first episode1
 - 45% after first recurrence²
 - -65% after two or more recurrences3
- Not a consequence of resistance to the 2 major treatment drugs: metronidazole or vancomycin
- Several empirical approaches to management have been advocated but most have no controlled data

Aslam S, et al. Lancet Infect Dis. 2005;5:549-557.
 McFarland LV, et al. Am J Gastroenterol. 2002;97:1769-177

23

Strategies Currently Available for Managing Recurrent CDI

- · Vancomycin* in tapering/pulsed doses
- · Saccharomyces boulardii
- Rifaximin (400 mg bid x 14d) following
 vancomycin* (the so called 'Chaser' regimen)
- Nitazoxanide (500 mg bid x 10d)
- IVIG (300 500 mg/kg)
- Fecal transplantation
- ??Fidaxomicin* (200 mg bid x 10d)

*Vancomycin & fidaxomicin are the only FDA-approved agents for CDI

Johnson S. J Infect. 2009;58:403-41

Vancomycin Taper/Pulse Regimen 125 mg QID 10–14 days 125 mg BID 7 days 125 mg daily 7 days 125 mg once every 2 days 8 days 125 mg once every 3 days 15 days Kyne L, Kelly CP, Gut. 2001;48:152 Tedesco FJ, et al. Am. J Gastroenierot. 1985;80:867-868.

25

Bacteriotherapy for Recurrent CDI:
Changes in Fecal Microbiome

Paters. Day 3

Paters. Day 3

Conc. Day 6

Conc. Day 7

Conc. Day 6

Conc. Day 7

Conc. Day 6

Conc. Day 7

Conc. Day 6

Conc. Day 7

Co

Conclusions

- Clostridium difficile infection continues to cause significant morbidity and mortality
- Understanding the pathophysiology (particularly, the role of toxins & immune response) may assist in the development of specific treatment modalities - Antitoxins, vaccines, specific antibodies
- Better patient assessment may assist in determination of level of risk and appropriate treatment in individuals with CDI & recurrent CDI
- Better understanding of the enteric microbiome may lead to better modalities of biologic therapy (synthetic stool?)

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Question 1

- Documented risk factors for developing CDI include all of the following except:
 - 1. Diagnosis of inflammatory bowel disease
 - 2. History of receiving chemotherapy
 - 3. Antimicrobial drug exposure ≤ 1 month
 - 4. Eating veal cutlets in Canada

2	8	

Question 2

- The ATLAS bedside CDI score includes all of the following except:
 - 1. Age
 - 2. Temperature
 - 3. Systolic blood pressure
 - 4. Leukocyte count

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Emerging CDI Treatment Options and Infection Control

Stuart Johnson, MD DTM&H Loyola University Medical Center Hines VA Hospital

Disclosures

- Consultant: Optimer, ViroPharma, Astellas, Pfizer, Cubist, and Bio-K+
- · Grants: VA Research Service
- FDA approved CDI Treatment Agents: Vancomycin, Fidaxomicin

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A patient sees you with frequent watery stools, tender abdomen and low grade fever, she just finished treatment for CDI seven days ago (metronidazole 500 mg tid for 14 days), her WBC is 12,000/mm³; you recommend:

- 1. Repeat metronidazole, same regimen
- 2. Repeat metronidazole, but treat for 3 weeks
- 3. Start vancomycin 125 mg qid for 10 days
- 4. Start nitazoxanide 500 mg bid for 10 days

3

Which of the following drugs does not have FDA approval for treatment of CDI?

- 1. Metronidazole
- 2. Vancomycin capsules
- 3. Vancomycin solution
- 4. Fidaxomicin

Overview:

- Current SHEA/IDSA treatment guidelines
- · Downsides of current CDI therapies
- Newly approved CDI treatment option
- · Current infection control strategies
- Potential future infection control strategies

5

Clinical Practice Guidelines for CDI in Adults: 2010 Update by SHEA/IDSA: Treatment Recommendations

- Metronidazole is the drug of choice for the initial episode of mild-moderate CDI (500 mg orally TID) for 10-14 days (A-I)
- Vancomycin is the drug of choice for an initial episode of severe CDI*. The dose is 125 mg orally QID for 10-14 days (B-I)
- · *Severe CDI defined as:
 - WBC > 15,000/mm3 or,
 - Cr > 1.5 x baseline

Cohen SH, et al. Infection Control Hospital Epidemiology. 2010;31:431-455.

Available Antibiotics for Treatment of CDI (prior to May 2011):

- Metronidazole
- Vancomycin (only FDA-approved agent for CDI prior to May 2011)
- Nitazoxanide
- Rifaximin
- Bacitracin
- Tigecycline (anectodal experience only for CDI)
- Teicoplanin**
- Fusidic Acid**

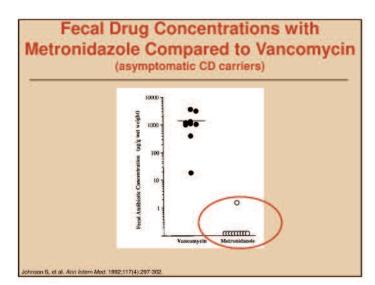
**Not available in the U.S.

Metronidazole

- · Widely used in the US after 1994 CDC HICPAC caution against PO vancomycin (due to concern about potential resistance in Enterococci)1
- · Decreased cure rates; slower response time compared to vancomycin²
- Risk of neurotoxicity with prolonged use³
- · Inferior to vancomycin for treatment of severe CDI⁴

Hospital Infection Control Practices Advisory Committee. MMWR Recomm Rep. 1995;44(RR-12):1-13. Others SA, et al. Infection Control and Hospital Epidemiology. 2010;31:431–455. Zar FA, et al. Dib Infect Dis 2007;45:302-307.

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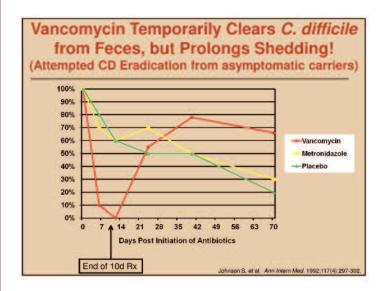


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Vancomycin

- Only agent approved for CDI in US by the FDA prior to May 2011
- · Highly effective, but:
 - Higher cost vs metronidazole
 - Potential for selection of vancomycinresistant Enterococci and Staphylococci
 - Frequent dosing
 - 20% (or more) rate of recurrence

Bauer, et al. Clin Microbiol Infect. 2009;15:1067–107.
 Kelly, LaMont. N Engl J Med. 2008;359:1932–1940.



11

Nitazoxanide (Prospective, Randomized, Comparative Trials for CDI)

- Compared to metronidazole (n=174)¹
 - Similar response rates, but also similar recurrence rates:

Antimicrobial	Response	Recurrence
Metronidazole 250 mg qid x 10d	28/34 (82.4%)	8/27 (29.6%)
Nitazoxanide 500 mg bid x 7d or 10d	68/76 (89.5%)	14/65 (21.5%)

- Compared to vancomycin (n=50)²
 - Similar response rates (94% vs 87%)
 - Study too small to confirm non-inferiority or to assess recurrence rates

Musher DM, et al. Clin Infect Dis. 2006;43:421–427.
 Musher DM, et al. Clin Infect Dis. 2009;48(4):e41–46.

1	2

Rifaximin

- Rifaximin "chaser" therapy for multiple recurrent CDI (400mg bid x 2 wks, immediately following last course of vancomycin)
 - No further recurrence: 7/8 pts
 - · 1 patient responded to 2nd rifaximin course
 - · Follow-up isolate was resistant to rifaximin
 - No further recurrence: 2/6 pts
 - · Rifaximin resistance identified in 1 isolate
- Resistance risk similar to other rifamycins
 - 36.8% of 470 recovered C. difficile isolates at 1 center were rifampin-resistant
- Johnson S, et al. Clin Infect Dis. 2007;44:846–848. Johnson S, et al. Anaerobe. 2009;15:290–291. Curry SR, et al. Clin Infect Dis. 2009;48:425–429.

- · Effective without recurrence
- Efficacious against virulent strains (eg, BI/027)

The "Ideal" CDI Treatment

- · Does not facilitate resistance
 - ... to *C. difficile* and other pathogens that reside in the gut, eg, *Enterococcus*
- Does not cross-react with clinically important antibiotics used for treating systemic infections eg, vancomycin
- Decreases spore shedding and transmission in the hospital setting
- · Good safety profile

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Newly Available CDI Therapy: Fidaxomicin

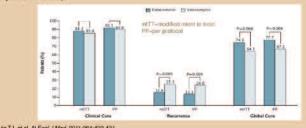
- Approved by FDA on May 27, 2011
- Indication and dosing¹
 - Treatment of Clostridium difficile-associated diarrhea (aka, CDI) in adults (≥18 years of age)
 - Recommended dose 200 mg orally bid for 10 d
- Advantageous characteristics^{1,2}
 - Minimal systemic absorption
 - Bactericidal agent unrelated to agents used for treatment of systemic infections
 - Narrow spectrum (less collateral damage to host flora)

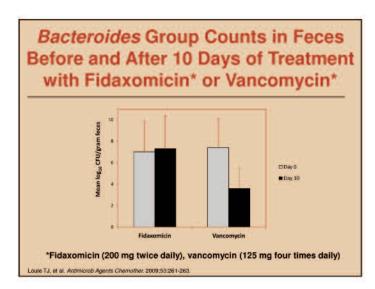
Dificid [package insert]. San Diego, CA: Optimer Pharmaceuticals; 2011

15

Fidaxomicin

- Rate of clinical cure with fidaxomicin non-inferior to that of vancomycin (phase 3 trial results)
- Fidaxomicin associated with significantly lower rate of CDI recurrence & similar adverse event profile
- Results of first phase 3 trial (nearly identical results from 2nd phase 3 trial):





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Cure Rates for Fidaxomicin & Vancomycin in CDI Patients infected with BI- and non-BI Strains P = 0.021 P < 0.001 100.0 86 6 90.0 80.0 70.0 60.0 50.0 30.0 20.0 10.0 0.0

Fidaxomicin, Challenges

- · Fidaxomicin challenges include:
 - Which patients should receive fidaxomicin treatment?
 - How will fidaxomicin fit in with the SHEA/IDSA guidelines for treatment based on severity?
 - Hospital formulary inclusion
 - Post approval monitoring for unanticipated side effects, evidence for resistance
- Additional potential antimicrobial agents in pipeline:
 - Ramoplanin, CB-183,315,... other

Dificid [package insert]. San Diego, CA: Optimer Pharmaceuticals; 2011. Tannock GW, et al. *Microbiology*, 2010;156(Pt11):3354-3359.

Which of the following infection control interventions for CDI is not based on clinical evidence?

- Glove use when caring for patients with CDI
- 2. Hand washing with soap & water after gloving when caring for patients with CDI
- 3. Terminal cleaning of CDI patient rooms with bleach
- 4. Antimicrobial use restriction

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Current Infection Control Strategies: Methods to Prevent Horizontal Transmission of *C. difficile*

Barrier Precautions:

- Gloves

- Handwashing

- Gowns

- Patient Cohorting

· Cleaning, Disinfection, Disposables

- Patient Rooms

- Endoscopes

- Rectal Thermometers

21

Proven
Probable
Untested
Probable

Clinical Efficacy

Probable Probable

Proven

22

Randomized Study of Gloves for CD Intervention

- Four wards randomized
- Intervention
 - Education: gloves when handling body substances (stool, blood, urine)
 - Gloves placed at bedside
- Significant reduction in CDI rate on glove wards

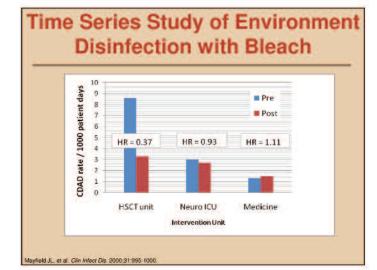
Johnson S, et al. Am J Med. 1990;88:137-140.

Hand Hygiene for CDI Intervention

- Appropriate hand hygiene area of controversy
 - In routine settings, alcohol-based hand hygiene in conjunction with isolation precautions using gloves may be acceptable
 - In setting of outbreak or increased rates, consider washing hands with <u>soap and water</u> after caring for patients with *C. difficile*

Cohen SH, et al. Infect Control Hosp Epidemiol. 2010;31:431-455.
 Dubberke Epi, et al. Infect Control Hosp Epidemiol. 2008;29:581-592.
 APIC Guide to the Elimination of Clastridium difficile in Healthcare Settings, Association for Professionals in Infection Control and Selectional Control and Selection Control and Selection Control Con





Current Infection Control Strategies: Methods to Reduce the Risk of *C. difficile* Diarrhea if Exposed to the Organism

Method Used

Efficacy Proven

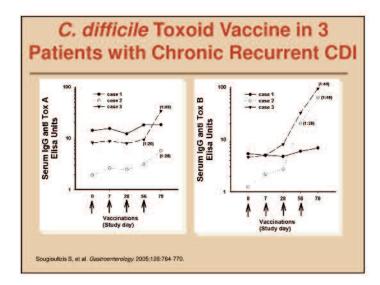
- Antimicrobial use restriction (clindamycin, 3rd gen cephalosporins, ?fluoroquinolones)
- Prophylactic treatment of patients receiving antimicrobials probiotics

Possible

Potential Future Strategies for Prevention

- · Primary Prevention
 - Active Vaccination
 - 'Effective Probiotics/Biotherapeutics'
- Secondary Prevention
 - Adjunctive Monoclonal Antibodies
 - Luminal Toxin Binders (tolevamer: No current plans for development)

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Provocative Probiotic Primary Prevention Studies for

Antibiotic-assoc Diarrhea (ADD) and C. difficile infection (CDI)

Probiotic Drink* Study

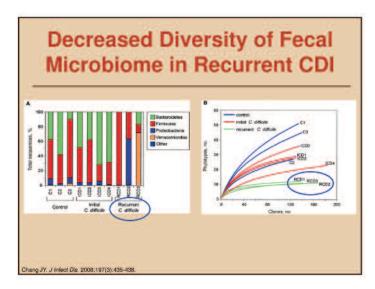
Placebo Probiotic
AAD 34% 12%
CDI 17% 0%

Probiotic Capsule† Dose-ranging Study

Placebo Probiotic-1 Probiotic-2
AAD 44% 28% 16%
CDI 24% 9% 1%

 Multiple criticisms of these studies; exclusion criteria, study blinding, & unexplained high rate of CDI in controls

Lactobacillus casei, L bulgaricus, Streptococcus thermophilus Lactobacillus acidophilus CL1285, Lactobacillus casei LBC80F

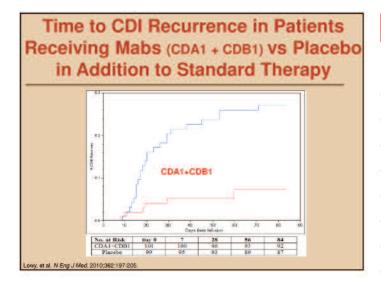


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Prevention of Fatal Infection with
Toxigenic C. difficile (J9) by Prior
Colonization of Hamsters with
Non-toxigenic C. difficile (M3)

Clindamycin
Day 1 Day 2 Day 3 Day 4 Day 5 Day 6 Day 7 Day 8 Day..

M3
J9
Control
J9
Control
X-dead



31

Conclusions:

- Currently available antimicrobial agents for treatment of CDI have several limitations; eg, recurrence after initially successful treatment
- Fidaxomicin, a newly-approved antimicrobial that appears to be a more narrow spectrum agent shows promise as an improved treatment for CDI
- Proven infection control interventions include gloving, antimicrobial restriction, and replacing rectal thermometers with disposable units; Hand hygiene & environmental decontamination with sporocidal agents are likely important, but may not be needed universally
- Future prevention strategies include immunotherapy & more effective probiotics

Clostridium	difficile	nfection:	Emerging	Therapies	and Re	currence	Prevention

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Deshpande A, Pasupuleti V, Rolston DD, et al. Diagnostic Accuracy of Real-time Polymerase Chain Reaction in Detection of *Clostridium difficile* in the Stool Samples of Patients With Suspected *Clostridium difficile* Infection: A Meta-Analysis. CID 53: e81-e90.

Gilligan PH. Is a two-step glutamate dehyrogenase antigencytotoxicity neutralization assay algorithm superior to the premier toxin A and B enzyme immunoassay for laboratory detection of *Clostridium difficile? J Clin Microbiol*. 2008;46(4):1523-1525.

Guerrero DM, Chou C, Jury LA, Nerandzic MM, Cadnum JC, Donskey CJ. Clinical and Infection Control Implications of *Clostridium difficile* Infection With Negative Enzyme Immunoassay for Toxin. *Clin Infect Dis.* 2011;53(3):287-290.

Kvach EJ, Ferguson D, Riska PF, Landry ML. Comparison of BD GeneOhm *Cdiff* real-time PCR assay with a two-step algorithm and a toxin A/B enzyme-linked immunosorbent assay for diagnosis of toxigenic *Clostridium difficile* infection. *J Clin Microbiol*. 2010;48(1):109-114.

Louie TJ, Miller MA, Mullane KM, et al. Fidaxomicin versus vancomycin for *Clostridium difficile* infection. *N Eng J Med*. 2011;364(5):422-431.

Luo RF, Banaei N. Is repeat PCR needed for diagnosis of *Clostridium difficile* infection? *J Clin Microbiol*. 2010;48(10):3738-3742.

Musher DM, Manhas A, Jain P, et al. Detection of *Clostridium difficile* toxin: comparison of enzyme immunoassay results with results obtained by cytotoxicity assay. *J Clin Microbiol*. 2007;45(8):2737-2739.

Norén T, Alriksson I, Andersson J, Akerlund T, Unemo M. Rapid and sensitive loop-mediated isothermal amplification test for *Clostridium difficile* detection challenges cytotoxin B cell test and culture as gold standard. *J Clin Microbiol*. 2011;49:710-711.

Novak-Weekley, Marlowe EM, Miller JM, et al. *Clostridium difficile* testing in the clinical laboratory by use of multiple testing algorithms. *J Clin Microbiol*. 2010;48(3):889-893.

Planche T, Aghaizu A, Holliman R, et al. Diagnosis of *Clostridium difficile* infection by toxin detection kits: a systematic review. *Lancet Infect Dis.* 2008;8(12):777-784.

Quinn CD, Sefers SE, Babiker W, et al. *C. Diff* Quik Chek complete enzyme immunoassay provides a reliable first-line method for detection of *Clostridium difficile* in stool specimens. *J Clin Microbiol*. 2010;48:603-605.

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Sloan LM, Duresko BJ, Gustafson DR, Rosenblatt JE. Comparison of real-time PCR for detection of the tcdC gene with four toxin immunoassays and culture in diagnosis of *Clostridium difficile* infection. *J Clin Microbiol*. 2008;46(6):1996-2001.

Stamper PD, Alcabasa R, Aird D, et al. Comparison of a commercial real-time PCR assay for tcdB detection to a cell culture cytotoxicity assay and toxigenic culture for direct detection of toxin-producing *Clostridium difficile* in clinical samples. *J Clin Microbiol*. 2009;47(2):373-378.

Swindells J, Brenwald N, Reading N, Oppenheim B. Evaluation of diagnostic tests for *Clostridium difficile* infection. *J Clin Microbiol*. 2010;48 (2):606-608.

Tenover FC, Novak-Weekley S, Woods CW, et al. Impact of strain type on detection of toxigenic *Clostridium difficile*: comparison of molecular diagnostic and enzyme immunoassay approaches. *J Clin Microbiol*. 2010;48(10):3719-3724.

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